

# Phase I Study of CBT -I™ and Taxol® in Patients with Taxol® Resistant Cancers

Robert K. Oldham, MD,<sup>1,2</sup> William K. Reid, MD,<sup>1</sup> and Daryl Barnett<sup>2</sup>

<sup>1</sup>Cancer Treatment Associates, Franklin, TN and <sup>2</sup>CBA Research, Inc., Lexington, KY

*CBT -I™, a natural product, was studied as an MDR modulator with Taxol® (135 mg/M) in an escalating dose Phase I clinical trial.*

*CBT -I™ was administered orally at doses from 300 mg/m<sup>2</sup> to 500 mg/m<sup>2</sup> daily x 7. The MTD was determined to be 500 mg/m<sup>2</sup> with moderate nausea and occasional emesis. Side effects were previously attributable to Taxol® rather than the study drug. A total of 18 patients were registered on study with only one patient determined to be intolerant of CBT -I™ due to nausea and emesis. In this Phase I study four patients (3 breast, 1 SCLC) remained stable for greater than two cycles of treatment. No complete or partial responses were seen in this Taxoflii resistant population.*

## INTRODUCTION

Although the biology and genetics of neoplastic disease are better understood and in spite of the introduction of many new treatments, cancer remains predominantly a progressive and fatal disease in its metastatic phase.

Chemotherapy is often initially successful in the treatment of patients with advanced solid tumors. Unfortunately, after a period of treatment, growth occurs as a result of acquired drug resistance leading to the subsequent death of the patient. In other solid tumors, de novo drug resistance is believed to be the cause of a lack of sensitivity to most chemotherapy agents. Thus, multi-drug resistance (MDR), whether inherent or acquired, appears to be the major failure mechanism for cancer chemotherapy in patients with advanced cancer.

While a variety of mechanisms of drug resistance are known or postulated, perhaps the best accepted mechanism involves the increased expression of the MDR1 gene which encodes the transmembrane glycoprotein Pgp. This mechanism appears to be the basis of resistance to multiple chemotherapy agents including *Vinca* alkaloids, anthracyclines, epipodophyllotoxins and taxanes. Pgp is a transmembrane glycoprotein that causes the active efflux of chemotherapy drugs in these classes with reduced intracellular accumulation of the drugs and subsequent drug resistance.<sup>1-9</sup>

---

*Address reprint requests to Robert K. Oldham, M.D.,  
Medical Director, Cancer Treatment Associates, P.O. Box  
680429, Franklin, TN 37068, Phone: 615/790-7535, Fax:  
615/794-9110.*

There is now evidence that MDR due to overexpression of Pgp can be reversed by several drugs, termed MDR modulators, some of which are now available for clinical use. Initial studies were done with calcium channel blockers such as verapamil.<sup>10</sup> Unfortunately, the plasma levels necessary to achieve reversal of MDR were excessively toxic in patients undergoing initial clinical trials with these agents.<sup>11</sup> Subsequently, studies with other Pgp modulators such as cyclosporine-A,<sup>12</sup> tamoxifen<sup>13</sup> and PSC-833<sup>14</sup> have been more encouraging.<sup>15-19</sup> In studies with cyclosporine-A and tamoxifen, MDR related to renal cancer could easily be reversed *in vitro*. Phase I trials determined the appropriate dose to be utilized with continuous infusion vinblastine and safely achieve plasma levels consistent with concentrations needed to achieve *in vitro* resistance reversal. Unfortunately, when a Phase II trial was done utilizing either high doses of cyclosporine-A or tamoxifen, no modulation of Inherent drug resistance was seen in patients with advanced renal carcinoma treated with continuous infusion vinblastine.<sup>20</sup> Pgp modulation was not measured and the authors felt the randomized trial design was cumbersome and inefficient. Since the modulation of clinical drug resistance is not likely to be useful in the clinic unless the effect is quite substantial, they suggested further Phase II trials with analyses referencing historical controls.<sup>21</sup>

More recent studies using cyclosporine<sup>22</sup> and cyclosporine derivatives such as PSC-833<sup>14</sup> have yielded interesting results. For example, in patients with advanced solid tumors treated with doxorubicin and PSC-833, an oral MDR modulator, a marked pharmacological interaction was noted between the chemotherapy drug and the MDR modulator. This led to significant hematological toxicity and required a reduction in the doxorubicin dose. A recent study of PSC-833 in acute myeloid leukemia utilized the modulator with reduced chemotherapy doses.<sup>23</sup> Although modulation was not measured in this study, plasma concentrations that could revert MDR *in vitro* were achieved in patients and further studies are planned with the suggestion that Pgp expression should be measured in the context of those studies.<sup>23</sup>

CBT-1<sup>TM</sup> is a natural product. While its mode of action has not been fully elucidated, experimental data with *in vitro* models demonstrated the drug's activity as an effective modulator of multiple drug resistance. CBT-1<sup>TM</sup> is felt to modulate Pgp expression, allowing for a greater intracellular accumulation of drugs and the reversal of drug resistance.<sup>24</sup>

This study was the second Phase I trial to define the tolerable dose range and side effects of CBT-1<sup>TM</sup> when administered with chemotherapy. The initial study was done with doxorubicin<sup>25</sup> and this study was conducted with Taxol®. In addition, pharmacokinetic studies were done of CBT-1<sup>TM</sup>, doxorubicin and Taxol® during the conduct of the initial trial.

## **PATIENTS AND METHODS**

This was a two-institution Phase I trial conducted with sponsorship of CBA Research, Inc.

### **Eligibility Criteria**

Patients with advanced solid tumors who were not curable by standard treatment and had failed a Taxol® containing regimen, were eligible. Patients were required to have a projected life

expectancy of > 12 weeks and a Kamofsky performance status of at least 60%. Normal liver and adequate kidney function (creatinine clearance  $\geq$  50cc per min), normal clotting and stable bone marrow function (white blood count  $\geq$  3000/mm<sup>3</sup>, platelet count  $\geq$  100,000/mm<sup>3</sup>) and normal EKGs were required. Women of childbearing age were excluded or had to be on an effective birth control method. Patients with significant coronary artery disease, cardiac arrhythmias or other active cardiac disease were not eligible. Patients were excluded if radiation therapy had been given within one week of entering study or if chemotherapy or other forms of systemic therapy were used within three weeks of entry. All of the patients were resistant to Taxol®. This study was registered with the Food and Drug Administration and had approval by the institutional review board in each participating institution. Each patient gave informed written consent prior to entering the study. Patient characteristics are shown in Table I.

## **Study Design**

In this Phase I study, CBT –1™ was to be administered to 18 patients by mouth on days 1-7 of each 21-day cycle in a dose escalating fashion. Taxol® was administered intravenously on day 6 of each cycle at a dose of 135 mg/m<sup>2</sup>. The dose of CBT –1™ was escalated by cohort with six patients to receive a dose of 300 mg/m<sup>2</sup>; seven patients 400 mg/m<sup>2</sup> and the final cohort of five patients to receive a dose of 500 mg/m<sup>2</sup>.

CBT –1™ was supplied by the sponsor, CBA Research Inc., as 50 mg capsules. These capsules were stored in the respective pharmacies at room temperature. Taxol® was supplied by each pharmacy. In selected patients, serum and urine CBT –1™ levels were periodically measured.

In this study, patients who were stable or responding at the end of cycle 2 were allowed to remain on treatment for up to six months from the start of therapy. Patients who developed a complete response within six months were to be treated with an additional two cycles and then therapy was to be discontinued. Tumor measurements were performed every two cycles and response definitions were standard with complete response being defined as disappearance of all clinically detectable disease, partial response defined as 50% or greater decrease in the sum of the products of two perpendicular diameters of all measurable disease without an increase in size of any single lesion or the appearance of any new lesions. Progressive disease was defined as greater than a 25% increase of the sum of the products of the two greatest perpendicular diameters or the appearance of new lesions. Stable disease was anything in between partial response and progressive disease. A response or stable disease was defined to exist if it persisted for at least two cycles. All registered patients, even if never treated, were included in this analysis. Patients were considered eligible for evaluation of response in this Phase I study if they completed two cycles of treatment.

## **Pretreatment Evaluation**

Prior to treatment, each patient was evaluated by physical examination and appropriate blood and urine tests. A full radiographic evaluation was conducted for measurement of disease and periodic chemical and radiological evaluations were performed to assess the toxicity of the regimen as well as to evaluate the patients for evidence of response.

## **Dose Modifications**

Patients with Grade 3 or 4 toxicity (standard ECOG toxicity criteria were used) had the next cycle delayed until signs and symptoms cleared. Patients with Grade 4 myelosuppression received the next dose of Taxol® at a 25% dose reduction. Patients with grade 3 or 4 toxicity thought to be due to CBT-1™ received their next dose at 100 mg/m<sup>2</sup> less of CBT-1™. All such CBT-1™ related toxicities were reported to the FDA by the sponsor .

## **Statistics**

CBT-1™ levels were performed by HPLC. As is documented in a previous paper, the assay is highly reproducible.<sup>25</sup> Data were analyzed by standard statistical tests calculating mean and standard deviation for each cohort of patients.

## **RESULTS**

The study was successfully conducted with 18 patients registered on the dose-escalating Phase I trial. Three (3) patients did not complete cycle 1 due to disease progression. Two (2) patients received a single cycle and 13 received 2 or more cycles. All cohorts were completed and a total of five patients were treated at the highest dose level of 500 mg/m<sup>2</sup> (Table 2). Generally speaking, the CBT-1™ was well tolerated and the toxicities observed were primarily related to Taxol®. Toxicity was evaluated in all patients receiving at least one dose of CBT-1™. Peripheral blood cytopenia, particularly neutropenia, was the most significant toxicity observed. These data are summarized in Table 3. Significant thrombocytopenia and anemia was seen occasionally.

Hemolysis studies were performed because of some previous evidence that CBT-1™ or its vehicle given intravenously could cause hemolysis. No change in serum haptoglobin or in urine hemosiderin levels were noted. ACTH and cortisol levels were generally within normal range on day 6. Kidney and liver function tests were normal throughout the study.

Gastrointestinal toxicity was felt to be drug related in one patient each receiving 300 mg/m<sup>2</sup> and 400 mg/m<sup>2</sup> CBT-1™. Of the 5 patients who did not complete the two cycles, 4 had nausea but all had disease complications felt to be the cause of their gastrointestinal signs. The 5 patients receiving 500 mg/m<sup>2</sup> had no complaints of nausea.

Because of the increasing frequency of nausea and occasional vomiting at doses of 600 mg/m<sup>2</sup> in the earlier Phase I study, the study was terminated at 500 mg/m<sup>2</sup> in this study.

## **Antitumor Activity**

Of the 18 patients entered in this Phase I study, 13 received at least 2 cycles of treatment and were evaluable for response assessment. Of those assessed there were 9 with progressive disease and 4 with stable disease. These data are summarized in Table 5.

## DISCUSSION

CBT-1™ plasma levels sufficient to reverse drug resistance in vitro were found at doses above 200 mg/m<sup>2</sup> in this study and the previous Phase I study.<sup>25</sup> It is clear from these studies that CBT-1™ can be administered orally and adequate plasma levels for MDR modulation are achieved. In addition, unlike cyclosporine and PSC-833, there was no significant alteration in the pharmacokinetics of Taxol® (unpublished data) and doxorubicin<sup>25</sup> or their toxicities when used with CBT-1™. Therefore, CBT-1™ appears to be an excellent drug for further investigation of MDR modulation.

The most frequent toxicity seen in this study was myelosuppression, which was felt to be Taxol® related. Mild nausea and some vomiting were seen at the 300 and 400 mg/m<sup>2</sup> dose levels of CBT-1™.

CBT-1™ hyperpigmentation was seen in several patients in this study but the other side effects noted were felt to be primarily related to Taxol® or the disease. The lack of significant pharmacological interaction between CBT-1™ and doxorubicin or Taxol® is very encouraging. Unlike verapamil, cyclosporine and PSC-833 where clear pharmacokinetic interactions affect the toxicity of chemotherapy, CBT-1™ can be administered in tolerable oral doses and plasma levels adequate to modulate MDR effects are achieved without significant changes in the pharmacokinetics of doxorubicin and Taxol®.<sup>14,26-28</sup> Studies of MDR-1 expression and Pgp levels were not done with this Phase I study but are planned in the future.

No neurological side effects were seen with CBT-1™, unlike studies with tamoxifen and PSC-833 where significant cerebellar ataxia has been observed.<sup>14,20,29,30</sup>

The antitumor activity seen in 4 of 13 patients with multidrug resistant disease is encouraging. Since oral CBT-1™ can be given safely with the usual therapeutic dose of doxorubicin and since CBT-1™ plasma levels achieved at tolerable doses are consistent with those that will modulate MDR in vitro, this drug appears to be a very interesting agent for Phase II/III evaluation.

Not only is it important to study MDR modulators in patients with resistant disease, it will also be important to combine CBT-1™ with chemotherapeutic agents prior to the development of resistance to determine whether they can increase the response rate to the chemotherapeutic agent and reduce the development of acquired drug resistance during the context of these trials.<sup>31</sup> Such studies may lead to improved response rates and survival times for patients with advanced cancer.

## ACKNOWLEDGMENTS

Technical assistance and CBT-1™/Taxol® assays and interpretation were provided by Hansun Ning, M.D., CBA Research. Dr. Oldham currently serves as Medical Director for CBA Research, Inc. Funding for this study was provided by the sponsor, CBA Research, Inc.

## REFERENCES

1. Bell DR, Gerlach JH, Kartner N, et al. Detection of P-glycoprotein in ovarian cancer. A molecular marker associated with multidrug resistance. *J Clin Oncol* 1985;3:311-315.
2. Pinedo HM and Giaccone G. P-glycoprotein: a marker of cancer cell behavior. *New Engl J Med* 1995;333: 1417-1419.
3. Beck W, Cirtain M, Danks M, Felsted R, Safa A, Wolverson J, Suttle D, and Trent J. Pharmacological, molecular and cytogenetic analysis of "atypical" multidrug-resistant human leukemic cells. *Cancer Res* 1987; 47:5455-5460.
4. Biedler J and Meyers M. M ultidrug resistance (*Vinca* alkaloids, actinomycin D, and anthracyclines antibiotics). In: R Gupta (ed.), *Drug Resistance in Mammalian Cells*, Vol. 2, Anticancer and Other Drugs, pp. 125-140, 1989. Boca Raton, FL: CRC Press.
5. Gerlach J, Kartner N, Bell D, and Ling V. Multidrug resistance. *Cancer Surv* 1986;5:25-46.
6. Fojo A, Ueda K, Slamon D, Poplack D, Gottesman M, and Pastan I. Expression of a multidrug-resistance gene in human tumors and tissues. *Proc Natl Acad Sci USA* 1987;84:265-269.
7. Yuen A and Sikic B. Multidrug resistance in lymphomas. *J Clin Oncol* 1994;12:2453-2459.
8. Fojo A, Shen D-W, Mickley L, Pastan I, and Gottesman M. Intrinsic drug resistance in human kidney cancer is associated with expression of a human multidrug resistance gene. *J Clin Oncol* 1987;5:1922-1927.
9. Goldstein I, Galski H, Fojo A, Willingham M, Lai S-L, Gazdar A, Pirker R, Green A, Crist W, Brodeur G, Lieber M, Cossman J, Gottesman M, and Pastan I. Expression of a multidrug resistance gene in human tumors. *J Natl Cancer Inst* 1989;81: 116-124.
10. Tsuruo T, Iida H, Tsukagoshi S, and Sakurai Y. Increased accumulation of vincristine and Adriamycin in drug resistant P388 tumor cells following incubation with calcium antagonists and calmodulin inhibitors. *Cancer Res* 1982;42:4730-4733.
11. Pennock G, Dalton W, Roeske W, Appleton C, Mosley K, Plezia P, Miller T, and Salmon S. Systemic toxic effects associated with high-dose verapamil infusion and chemotherapy administration. *J Natl Cancer Inst* 1991;83:105-110.
12. Samuels B, Mick R, Vogelzang N, Williams S, Schilsky R, Safa A, O'Brien S, and Ratain M. Modulation of vinblastine resistance with cyclosporine: a Phase I study. *Clin Pharmacol Ther* 1993;54:421-429.
13. Trump D, Smith D, Ellis P, Rogers M, Schold S, Winer E, Panella T, Jordan V, and Fine R. High-dose oral tamoxifen, a potential multidrug -resistance- reversal agent: Phase I trial in combination with vinblastine. *J Natl Cancer Inst* 1992;84:1811-1816.
14. Giaccone G, Linn SC, Welink J, Catimel G, Steltjes H, van der Vijgh WJF, Eelink C, Vermorken JB, and Pinedo HM. A dose-finding and pharmacokinetic study of reversal of multidrug resistance with SDZ PSC 833 in combination with doxorubicin in patients with solid tumors. *Clin Cancer Res* 1997;3:2005-2015.
15. Leith CP, Chen I-M, Kopecky KJ, et al. Correlation of multidrug resistance (MDR1) protein expression with functional dye/drug efflux in acute myeloid leukemia by multiparameter flow cytometry: Identification of discordant MDR-/efflux+ and MDR1 +/efflux- cases. *Blood* 1995;86:2329-2342.

16. Solary E, Witz B, Caillot D, et al. Combination of quinine as a potential reversing agent with mitoxantrone and cytarabine for the treatment of acute leukemias: A randomized multicenter study. *Blood* 1996;88: 1198-1205.
17. Miller TP, Grogan TM, Dalton WS, et al. P-glycoprotein expression in malignant lymphoma and reversal of clinical drug resistance with chemotherapy plus high dose verapamil. *J Clin Oncol* 1991;9:17-24.
18. Ross DD, WootenPJ, Sridhara R, et al. Enhancement of daunorubicin accumulation, retention, and cytotoxicity by verapamil or cyclosporin A in blast cells from patients with previously untreated acute myeloid leukemia. *Blood* 1993;82:1288-1299.
19. List AF , Spier C, Greer J, et al. Phase I/II trial of cyclosporine as a chemotherapy-resistance modifier in acute leukemia. *J Clin Oncol* 1993;11:1652-1660.
20. Samuels BL, Hollis DR, Rosner GL, Trump DL, Shapiro CL, Vogelzang NJ, and Schilsky RL. Modulation of vinblastine resistance in metastatic renal cell carcinoma with cyclosporine A or tamoxifen: a cancer and leukemia group B study. *Clin Cancer Res* 1997;3:1977-1984.
21. Simon R. Designs for efficient clinical trials. *Oncology* 1989;3:43-49.
22. Bartlett NL, Lum BL, Fisher GA, Brophy NA, Ehsan MN, Halsey J, and Sikic IB. Phase I trial of doxorubicin with cyclosporine as a modulator of multidrug resistance. *Journ Clin Oncol* 1994;12:835-842.
23. Lee EJ, George SL, Caligiuri M, et al. Parallel phase I studies of daunorubicin given with cytarabine and etoposide with or without the multidrug resistance modulator PSC-833 in previously untreated patients 60 years of age or older with acute myeloid leukemia: results of cancer and leukemia group B study 9420. *J Clin Oncol* 1999;17(9):2831-
24. CBA Research Inc. Data on file, 1998.
25. Oldham RK, Reid WK, Preisler HD, and Bamett D. A phase I and pharmacokinetic study of CBT -1 as a multidrug resistance modulator in the treatment of patients with advanced cancer. *Cancer Biotherapy & Radiopharmaceuticals* 1998;13(2):71-80.
26. Kerr DJ, Graham J, Cwnmings J, Morrison JG, Thompson 00, Brodie MJ, and Kaye SB. The effect of verapamil on the pharmacokinetics of adriamycin. *Cancer Chemother Pharmacol* 1986; 18:239-242.
27. Lum BL, Kaubish S, Yahanda AM, Adler KM, Jew L, Ehsan MN", Brophy NA, Halsey J, Gosland MP , and Sikic BI. Alteration of etoposide pharmacokinetics and pharmacodynamics by cyclosporine in a phase I trial to modulate Multidrug resistance. *J Clin Oncol* 1992;10: 1635-1642.
28. Speeg KV and Maldonado AL. Effect of the nonimmunosuppressive cyclosporine analog SDZ PSC-833 on colchicine and doxorubicin biliary secretion by the rat in vivo. *Cancer Chemother Pharmacol* 1994;34:133-136.
29. Boote DJ, Dennis IF, Twentyman PR, Osborne RJ, Laburte C, Hensel S, Smyth JF , Brampton MH, and Bleehen NM. Phase I study of etoposide with SDZ PSC-833 as a modulator of multidrug resistance in patients with cancer. *J Clin Oncol* 1996;14:610-618.
30. Fisher GA, Lum BL, Hausdorff J, And Sikic BI. Pharmacological considerations in the modulation of multidrug resistance. *Eur J Cancer* 1996;32A: 1082-1088.
31. Beketic-Oreskovic L, Duran, Chen G, et al. Decreased mutation rate for cellular resistance to doxorubicin and suppression of mdr 1 gene activation by the cyclosporin analogue PSC 833. *J Natl Cancer Inst* 1995;87:1593-1602.